

APRIL 5-10 #AACR24 AACR.ORG/AACR24



Ultra-sensitive ctDNA detection predicts response to immune checkpoint inhibition in advanced melanoma patients

Christoffer Gebhardt

University Medical Center Hamburg-Eppendorf (UKE)

Department of Dermatology and Venereology, University Skin Cancer Center Hamburg











Hubertus Wald Tumorzentrum Universitäres Cancer Center Hamburg

Ein Kompetenznetzwerk des UKE





Prof. Dr. med. Christoffer Gebhardt

Advisory Roles for, and Honoraria as well as Travel Expenses by the following Companies:

Almirall MSD Sharp & Dohme

Amgen Novartis

Beiersdorf Pierre-Fabre

BioNTech Regeneron

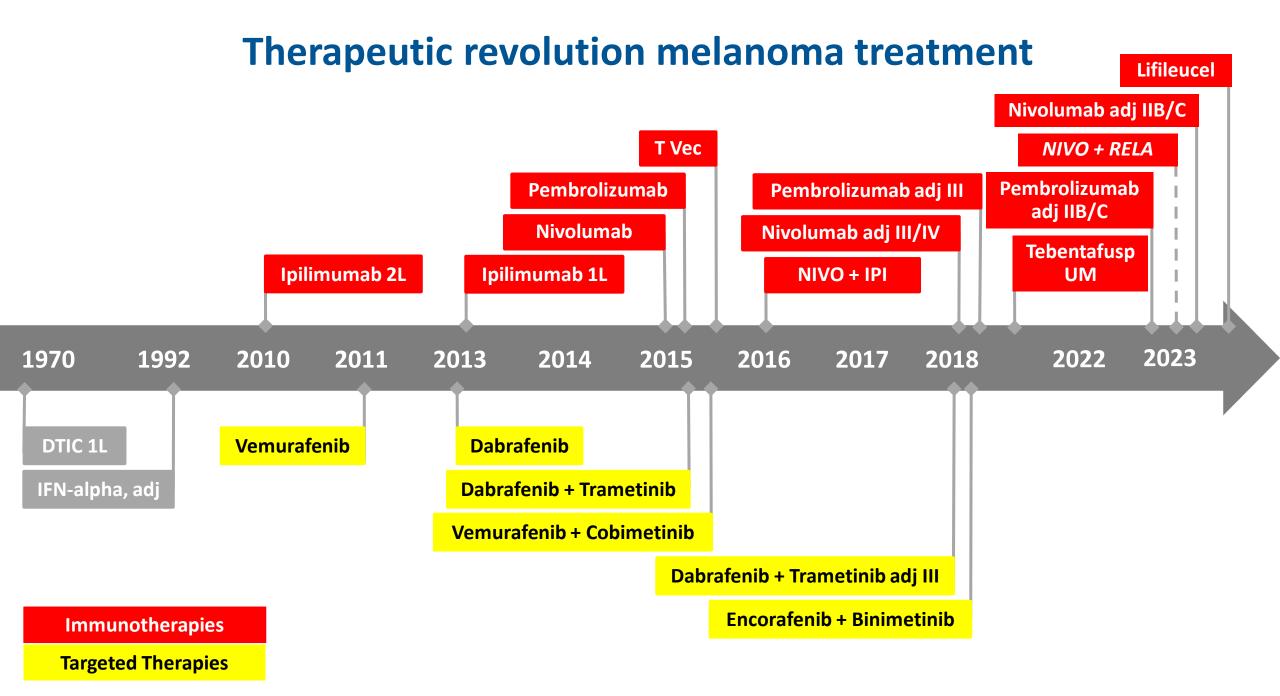
Bristol-Myers Squibb Sanofi Genzyme

Delcath Sciomics

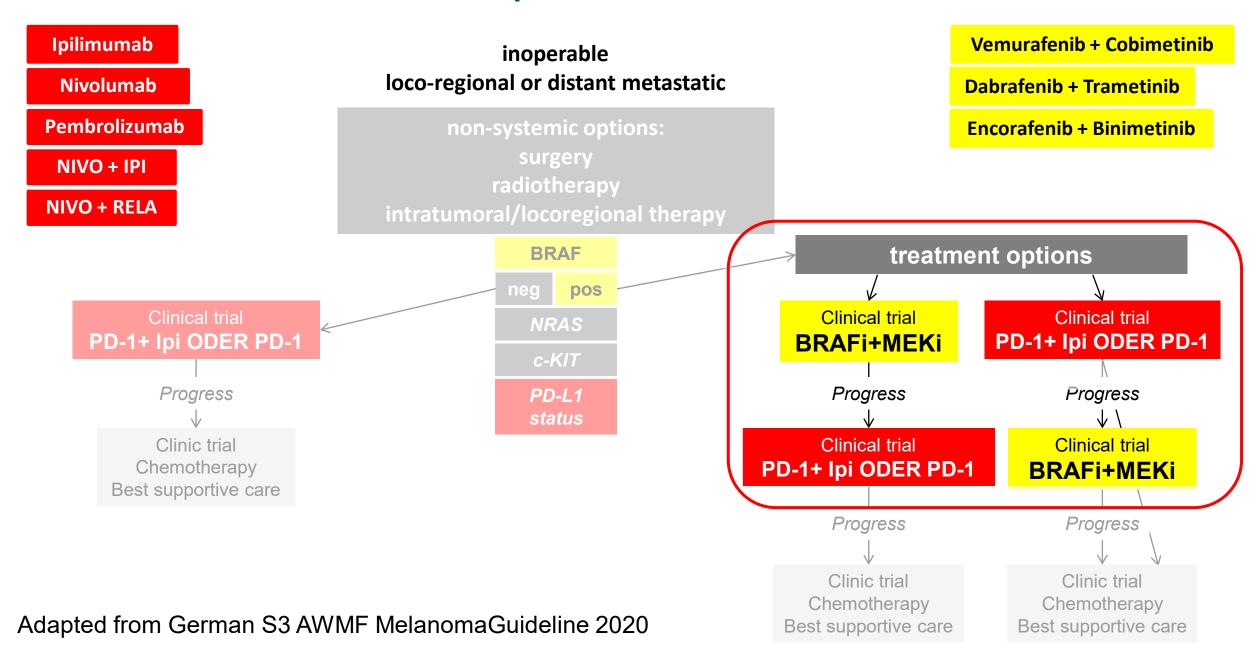
Immunocore SUN Pharma

Janssen Sysmex/Inostix

Co-Founder of Dermagnostix and Dermagnostix R&D



What is the best treatment sequence in metastatic BRAF mutant melanoma?



Biomarkers for therapy management in melanoma

Tissue-based (Tumor/Tumormicromilieu)

Immunoscore/

Immuneprofiling

PD-L1

Expression

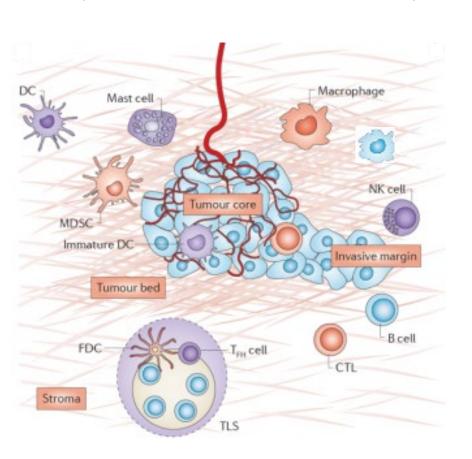
Gene

Expression

Signatures

IFN-gamma Signature

Tumor mutation load (TMB)



Fecal microbiome

Blood-based



S100B

LDH

ctDNA

(=cell-free Tumor-DNA)

CTCs

Chemokines/ Cytokines

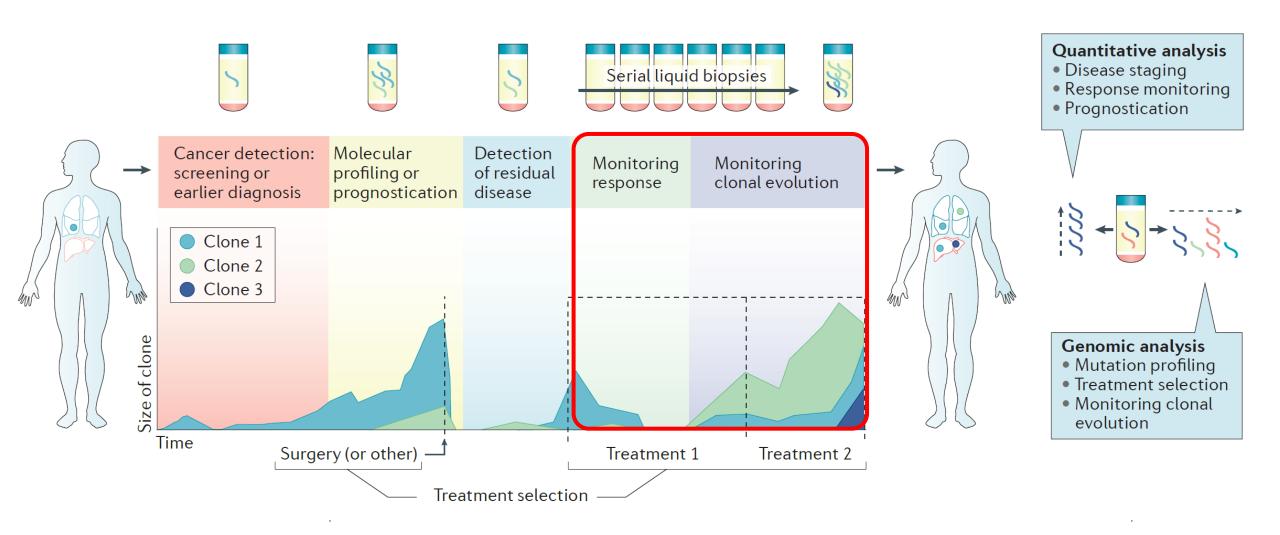
Exosomes

Immune cellsubtypes

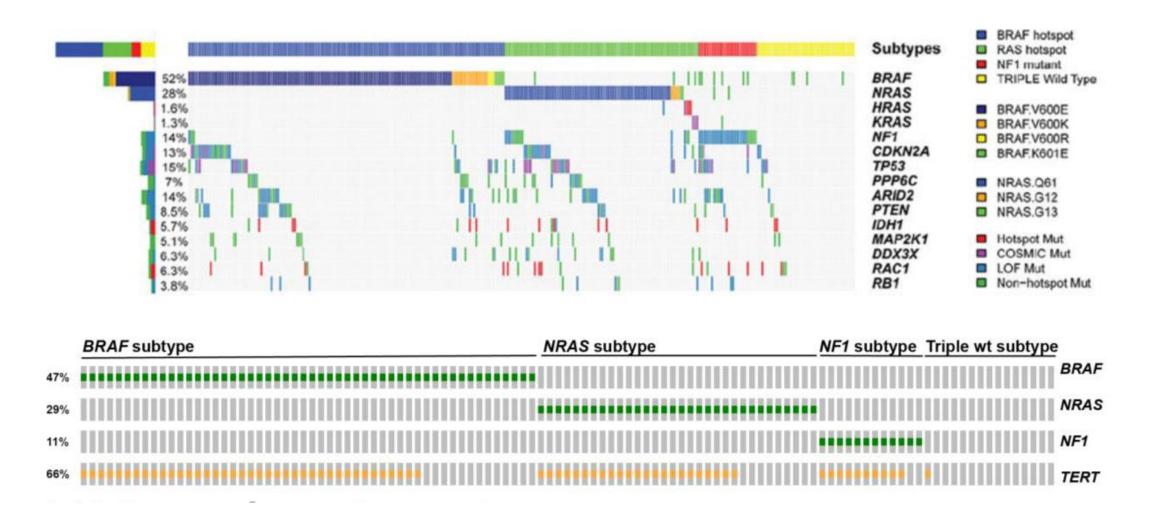
Metabolites

Clinical Biomarker

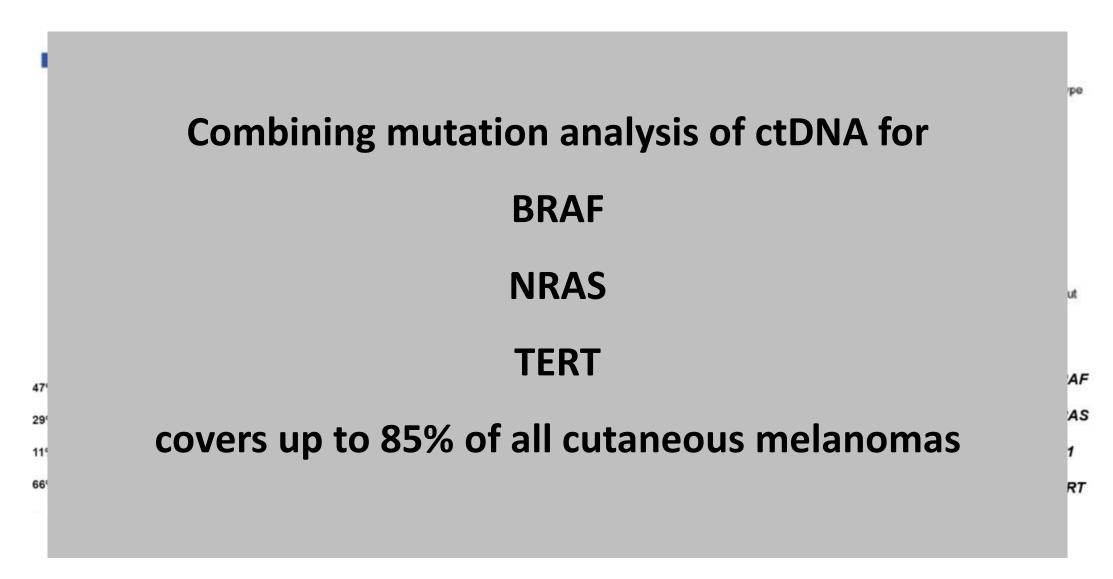
ctDNA diagnostics could guide sequencing of treatment



Driver Mutations in Melanoma: BRAF... and more



Driver Mutations in Melanoma: BRAF... and more

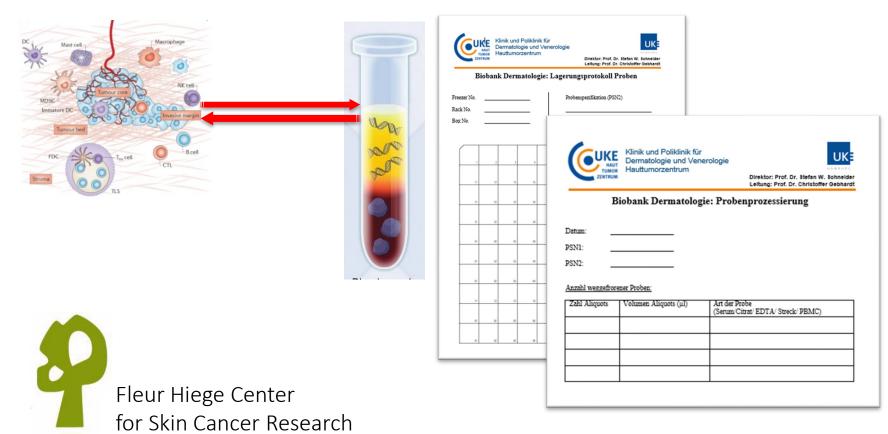




Establishment of the LiquiMEL Biobank

Multicenter Biobank together with Buxtehude, UKSH Lübeck and Kiel (since 2018)

To date >16,000 blood specimens of skin cancers patients receiving systemic treatment



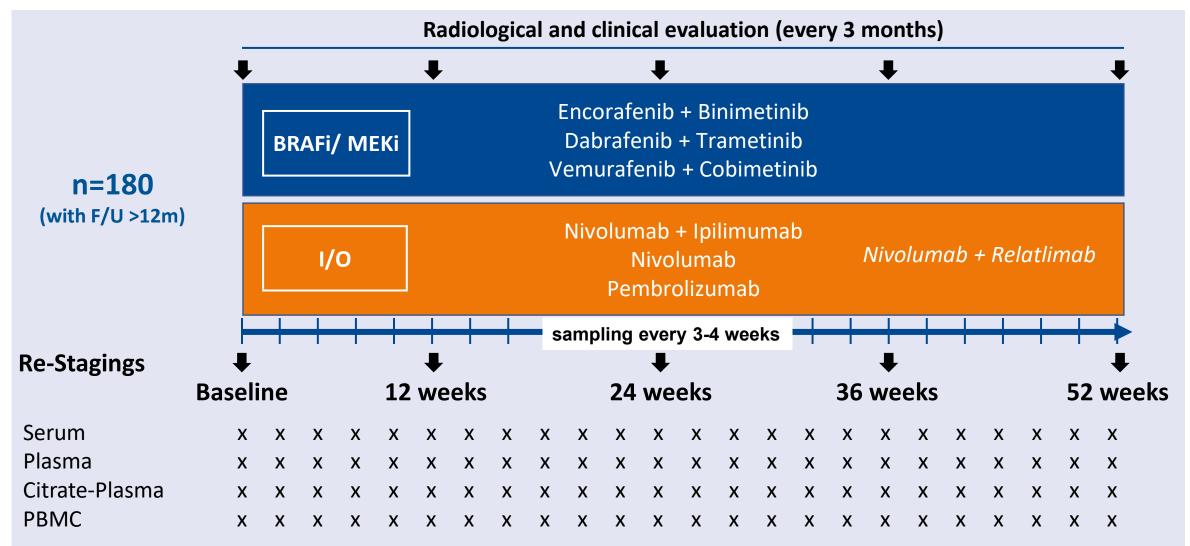
Collaborative Effort of Skin Cancer Center and Institute of Tumor Biology (Prof. Klaus Pantel)

Several collaborations internally, nationally and internationally

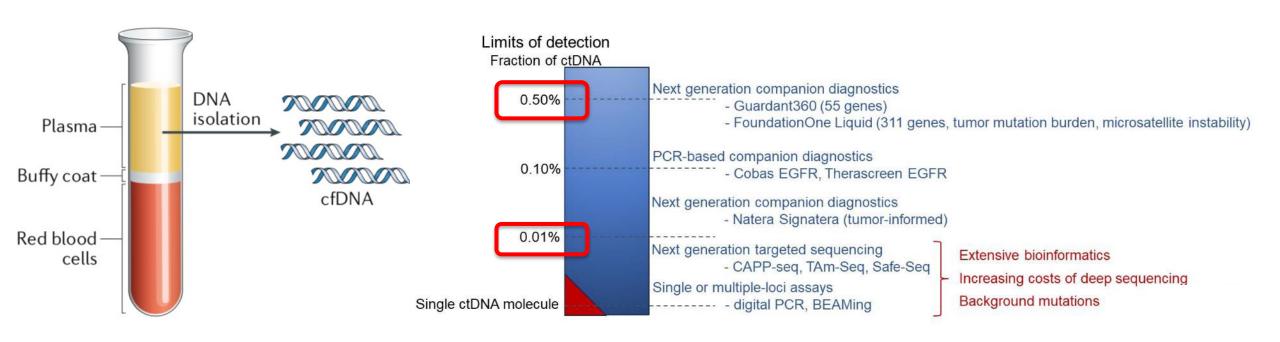


LiquiMEL Biobank

>230 non-resectable Stage III/IV Melanoma Patients (since 2018)

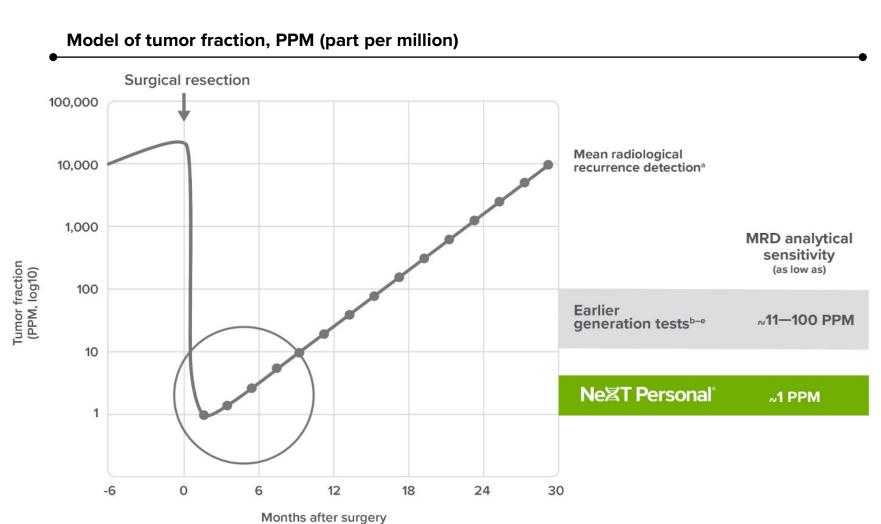


Methods of ctDNA Analysis and Limits of Detection

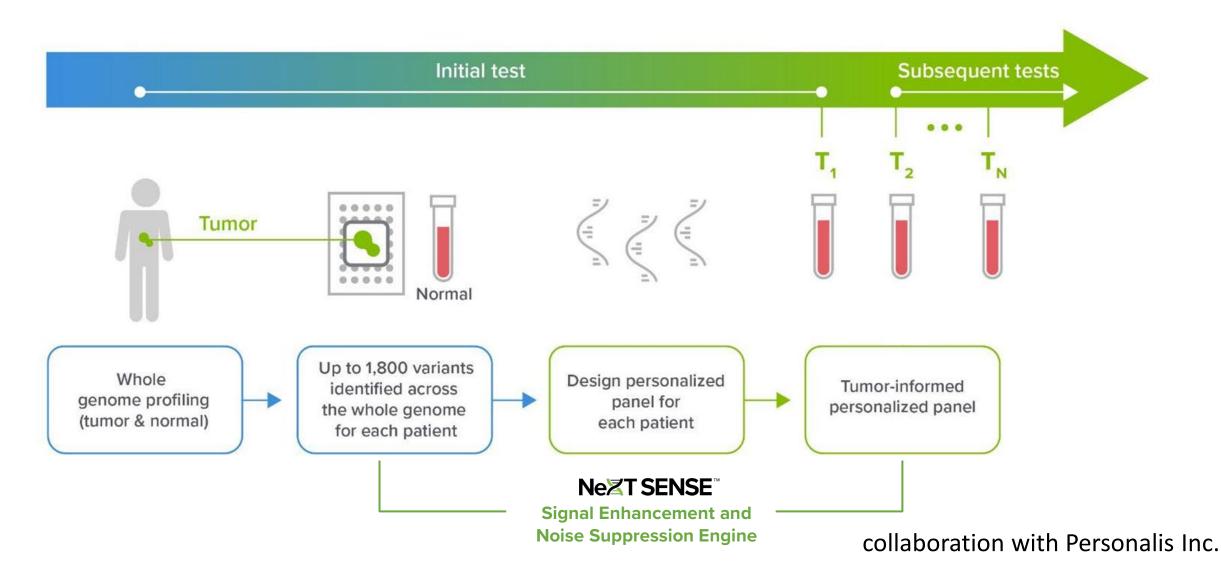


Standard ddPCR reaches sensitivity of up to 100 PPM

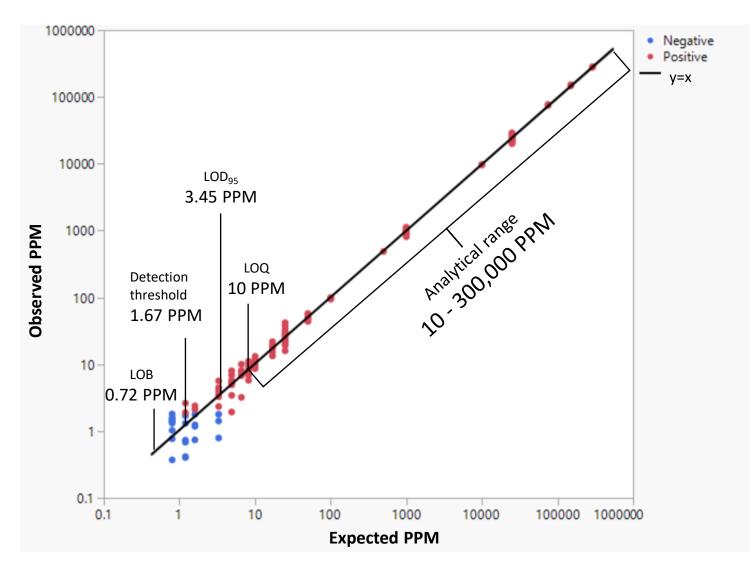
NeXT Personal is an ultra-sensitive and specific MRD test designed to see MRD earlier and more accurately



Tumor informed MRD approach powered by whole genome sequencing and advanced analytics



Analytical range of the NeXT Personal platform



Limit of blank (LOB): 0.72 PPM

The level of signal (noise) detected in instances where no tumor is present.

Detection threshold: 1.67 PPM in AV study

The lower limit at which a positive call can be made at the defined specificity; defines LOD_{50} .

95% Limit of detection (LOD₉₅): 3.45 PPM

The concentration at which 95% of reading would be positively detected.

Limit of quantification (LOQ): 10 PPM

The lower limit at which two measurements can be quantitatively distinguished.

Analytical range: 10 PPM - 300,000 PPM

The range over which which two measurements can be quantitatively distinguished.

Patient characteristics

Age at diagnosis	Median (SD)	55 (5.3)
Gender		
	F	8 (35)
	M	15 (65)

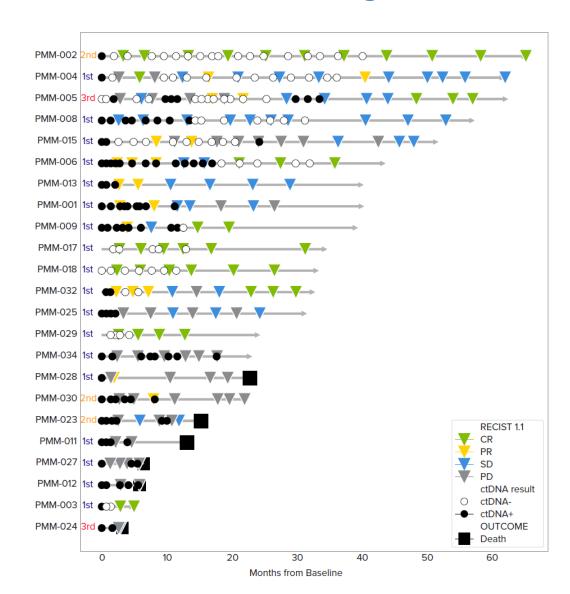
Melanoma subtype		
	Cutaneous	18 (78)
	Mucosal	3 (13)
	CUP	2 (9)

~		
S100B		
	Elevated	12 (52)
	Normal	11(48)
LDH		
	Elevated	16 (70)
	Normal	7 (30)
BRAF		
	wt	11 (48)
	mut	12 (52)

Tumor stage (AJCC version 8)		
	III	2 (9)
	IV	21 (91)
ICI Treatment		
	Pembro	3 (13)
	Nivo	2 (9)
	Ipi+ Nivo	18 (78)
Therapy line		
	1st	18 (78)
	2nd	3 (13)
	3rd	2 (9)
Best overall respon	nse to ICI	
	Complete resp	onse 10 (43.
	Partial respons	se 5 (21.7
	Stable disease	4 (17.4
	Progressive di	sease 4 (17.4)

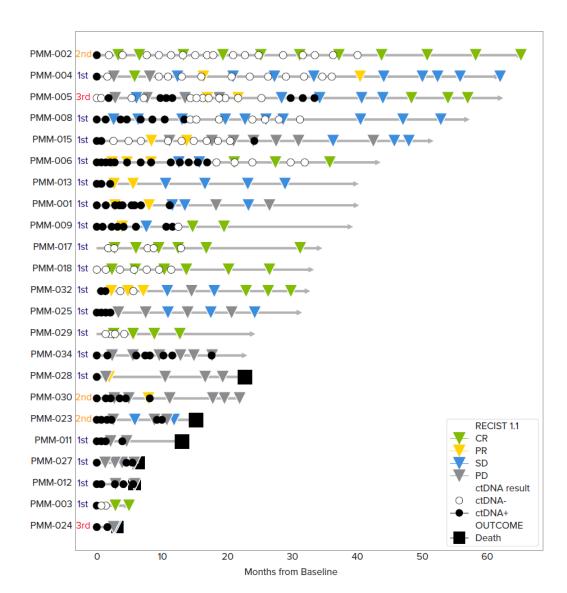
	None	17 (74)
	Present	7 (26)
Liver metastasis	(at treatment start)
	None	17 (74)
	Present	6 (26)
Lung metastasis	(at treatment start)
	None	8 (35)
	Present	15 (65)
Bone metastasis ((at treatment start))
	None	21 (91)
	Present	2 (9)

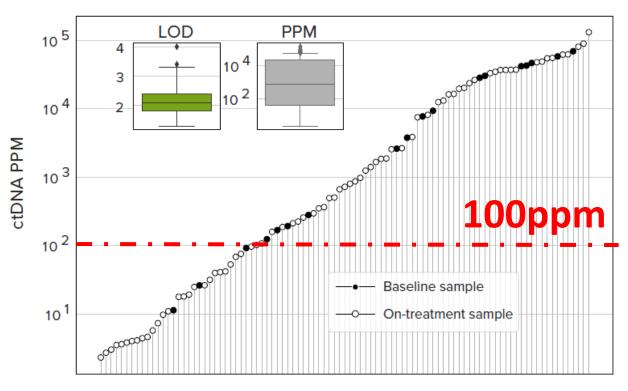
Majority of patient baseline samples detected using an ultra-sensitive ctDNA assay



188 plasma samples from 23 melanoma patients receiving ICI over several years

Majority of patient baseline samples detected using an ultra-sensitive ctDNA assay

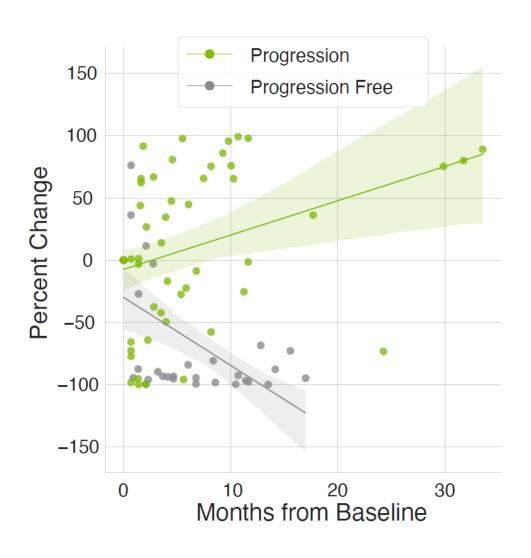


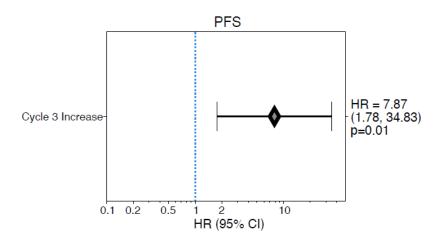


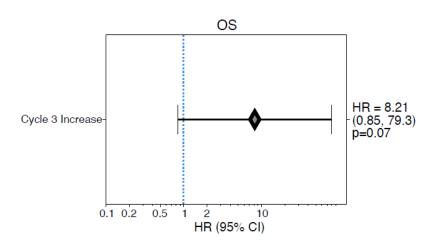
ctDNA was detected with a **sensitivity down to 2.3 PPM Up to 28.6% of all positive detections fall below 100 PPM**

Standard ddPCR reaches sensitivity of up to 100 PPM

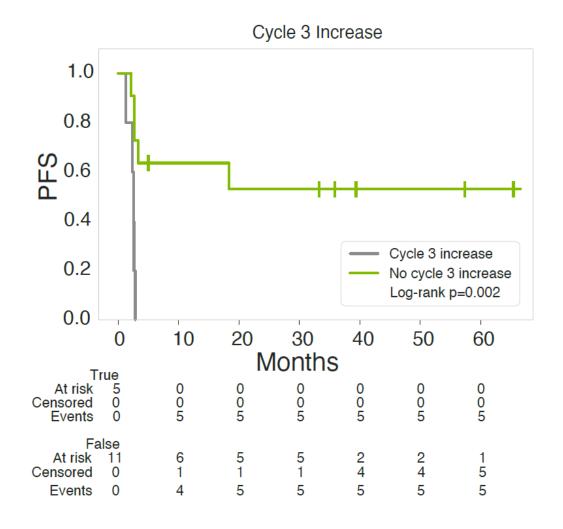
Early ctDNA increases from baseline are prognostic of progression-free survival

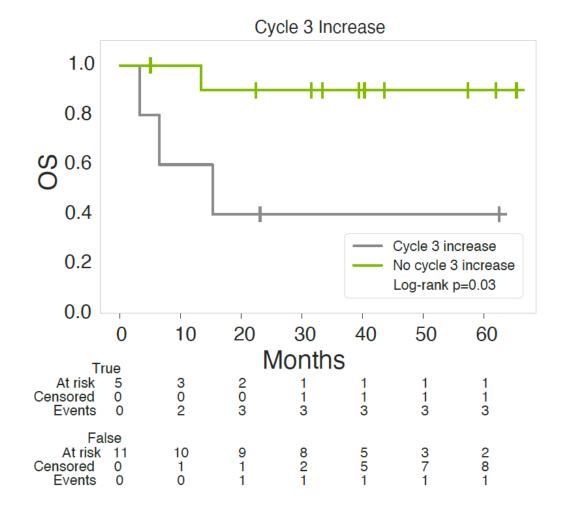




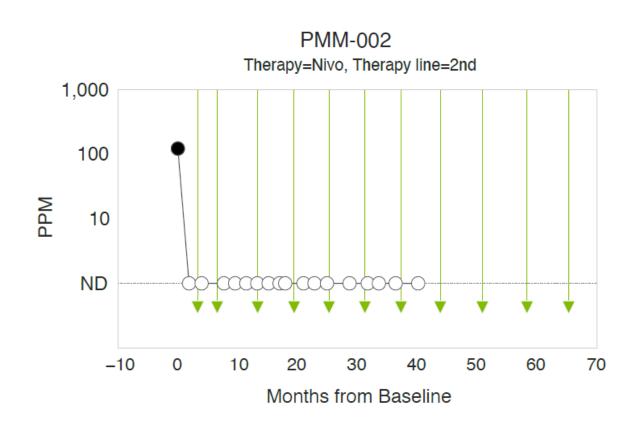


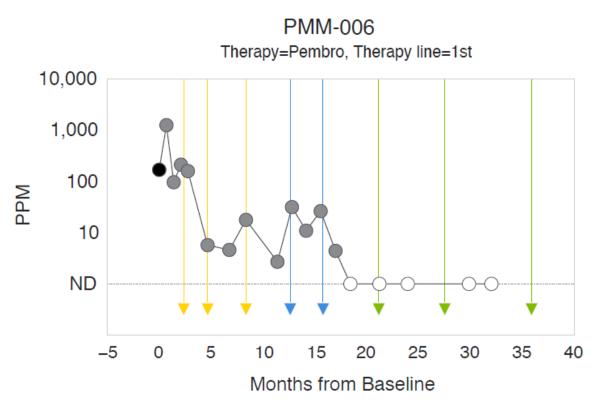
Early ctDNA increases from baseline are prognostic of progression-free survival





On-treatment ctDNA measures correlate with response and are prognostic of overall survival

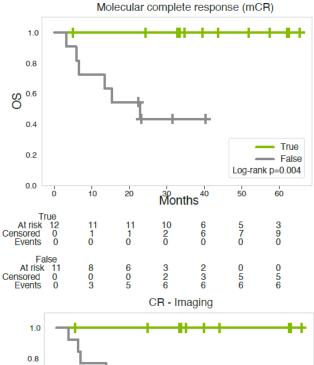




On-treatment ctDNA measures correlate with response and are prognostic of overall survival

Molecular Complete Response (mCR)

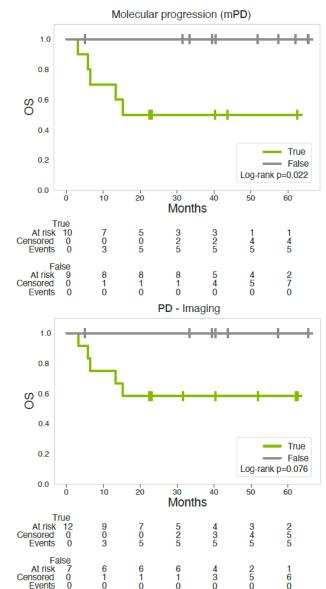
Radiological
Complete Response
(CR)



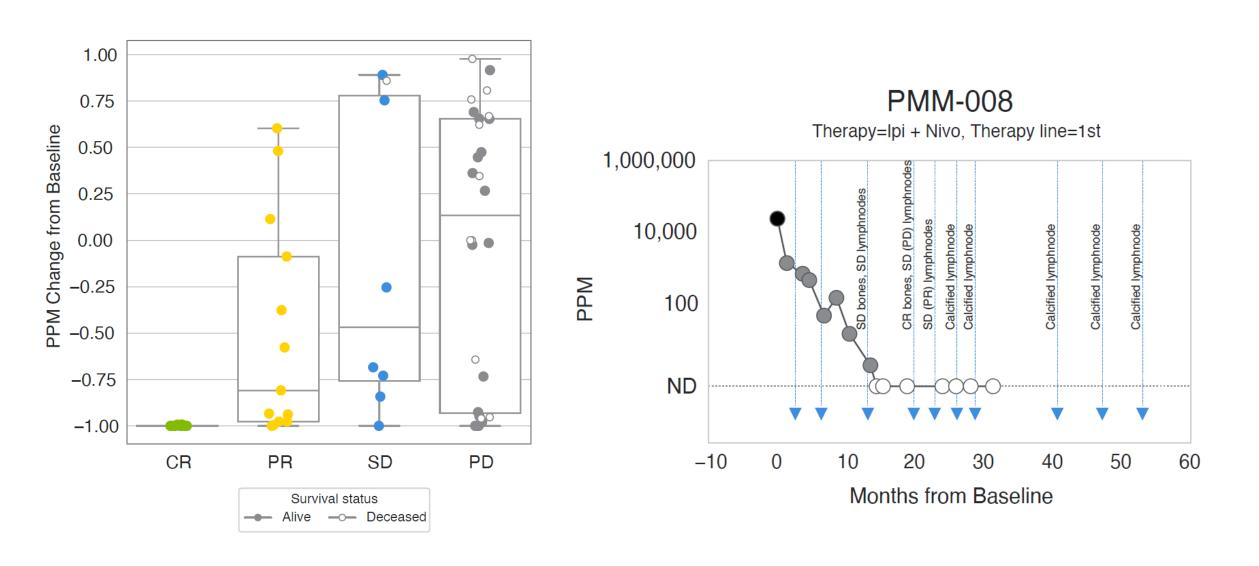
Events

Molecular Progressive Disease (mPD)

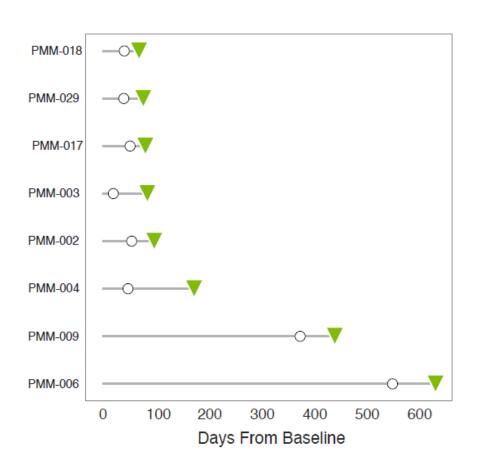
Radiological
Progressive Disease
(PD)



Pairing ctDNA and imaging-based response evaluation improves the accuracy of response classifications



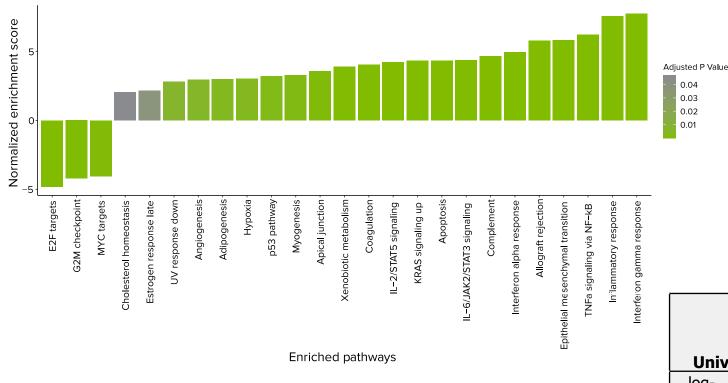
ctDNA status identifies response prior to imaging



Radiological CR was preceded by at least one ontreatment ctDNA negative determination in 100% (10/10) of CR patients,

with mCR occurring an average of 250 days prior to imaging based CR (median = 66 days)

Deep ctDNA monitoring and tumor immune profiling accurately predicts ICI resistance



NeXT Liquid Biopsy® (NeXT LB) platform

Multivariable Cox models of progression free survival that combine immune features with ctDNA measures compared to univariable assessment

Log-likelihood ratio Multivariable model test between models adjusted for ctDNA with and without **Univariable model** burden **ctDNA** logloglikelihoo p-value HR likelihoo p-value HR p-value Myeloid DC's 4.0533 0.0441 0.163 6.6057 0.0368 0.25 0.024 Neoantigen 9.6178 0.0019 0.17 11.6453 0.003 0.18 0.044 Burden 3.9206 CD8 T cells 0.0477 < 0.01 6.4836 0.0391 < 0.01 0.024

Wald; adjusted p<0.01), as measured by baseline RNA expression profiling

Abbott et al. AACR 2024, Poster #2415

Summary and Conclusions

- Despite remarkable progress in the treatment of melanoma by immune checkpoint inhibitors (ICI), a
 large proportion of patients still do not respond or achieve durable clinical benefit from ICI
- Circulating tumor DNA (ctDNA) monitoring in peripheral blood has shown promise for disease surveillance and prognostication of ICI response. Yet even in the metastatic context, ctDNA signals can be missed, suggesting the need for increased assay sensitivity
- We used a tumor-informed ctDNA assay capable of detection down to 1 part-per-million (PPM) ctDNA to profile 188 plasma samples from 23 melanoma patients receiving ICI over several years
- Residual disease was detected in ctDNA over a broad range, from 130,000 PPM down to 2.3
 PPM, with approximately a third below 100 PPM and a quarter of detections below 50 PPM
- Early on-treatment increases in ctDNA level, and both ctDNA-based molecular clearance (mCR) and molecular progression (mPD) were all prognostic of both OS and PFS
- Additionally, mCR consistently preceded radiographic imaging
- Our results show the importance and utility of an ultra-sensitive ctDNA assay in advanced melanoma treated by ICI with potential implications for other cancers



Contributors and Funding



Ein Kompetenznetzwerk des UKE

Dermatology, Essen

Dirk Schadendorf Alpaslan Tasdogan Selma Ugurel Alexander Rösch Antje Sucker

Dermatooncology, DKFZ

Jochen Utikal Claudia Czerwinska

Clinical Chemistry, Mannheim

Michael Neumaier Verena Haselmann

Biostatistics, DKFZ

Tim Holland-Letz

Dermatology, UKE
Stefan W. Schneider
Glenn Geidel
Julian Kött
Isabel Heidrich
Alessandra Rünger
Julia Stadler
Christian Gorzelanny
Alexander Bauer
Christian Mess

Laura Adam

Nadine Ammann

Chiara Blomen

Benjamin Deitert

Julia Gerdsen

Leah Goerdt

Inka Hoehne

Wieland Löffel

Myriam Merkle

Niousha Parnian

Charlotte Rautmann

Carmen Röper

Noemi Schlepper

Shari Schneider

Yasmin Fede Schwietzer

Mark Sementsov

Marie Steger

Carlotta Stramaglia

Tim Zell

Noah Zimmermann

Ohne Forschung

keine Zukunft

HAUTKREBSSTIFTUNG

Institute for Tumor Biology, UKE
Klaus Pantel

Laura Keller

Daniel Smit

Anatomy, UKE

Udo Schumacher

Oncology, UKSH Lübeck

Nikolas v. Bubnoff

Eva Danzer

Dermatology Buxtehude

Peter Mohr

Dermatology, UKSH Kiel

Axel Hauschild

Katharina Kähler

Dermatology, UKSH Lübeck

Patrick Terheyden





Fleur Hiege-Center for Skin Cancer Research



Fleur-Mareen Habig, née Hiege (1972-2005)



Collaborative research center of **University Skin Cancer Center**, Department of Dermatology and **Institute of Tumor Biology** at the University Medical Center Hamburg-Eppendorf (UKE)

Our Clinician Scientists in Skin Cancer Research:



Glenn Geidel



Isabel Heidrich



Julian Kött



Alessandra Rünger



Department of Dermatology and Venereology University Skin Cancer Center



















Martinistraße 52 | D-20246 Hamburg

Univ.-Prof. Dr. med. Christoffer Gebhardt

Stellvertretender Klinikdirektor und Leiter Hauttumorzentrum

Telefon +49 (0) 40 7410-57626

Telefax +49 (0) 40 7410-57545

ch.gebhardt@uke.de | www.uke.de

